Cytotoxicity and Inflammatory Effect of Silver Nanoparticles in Human Cells

Jeong-shin Park, Na Mi Yu, Jinwoo Cheon and In-Hong Choi

Department of Microbiology, College of Medicine; Department of Chemistry; Nanomedical NCRC, Yonsei University, Seoul, Korea 1. Approaches to practical toxicology tests to assess nanoparticles

2. Cytotoxicity and inflammatory effects of silver nanoparticles

Nanoparticles and toxicity assay

- The rapidly developing field of nanotechnology will result in exposure of nanoparticles to humans via several routes (e.g., inhalation, ingestion, skin, etc.). Nanoparticles can translocate from the route of exposure to other vital organs and penetrate cells.
- Toxicity studies to determine the deleterious effects of nanoparticles on living cells are required.
- Due to the nanosize and the nature of agglomeration, simple standard methods to characterize the biological effects of nanoparticles are currently unavailable.
- In this study, practical information regarding the optimal *in vitro* tests for nanotoxicity were evaluated.

Silver nanoparticles

Antimicrobial reagents, detergents, water purificants, wall paints, textiles



Biological tests





In vitro tests for nanoparticles



Exposure routes of nanomaterials



| | Cell line | Origin | Characteristics | |
|-------------|-----------|----------------------|---|--|
| Respiratory | A549 | Lung epithelial | Proper for cytotoxicity | |
| | BEAS-2B | Bronchial epithelial | Proper for cytokine | |
| Immune | U937 | Macrophage | Proper for cytotoxicity and cytokine production | |
| Skin | SK-Mel | Skin epithelial | Proper for cytotoxicity and cytokine production | |
| | A375 | Skin epithelial | Too fast growing | |

Standard toxicology tests and silver nanoparticles

| Category | Tests | Mechanism | Method | Suggestion |
|----------------------------------|---------------------------------|---|----------|---|
| In Vitro Immunology | Hemolysis | Release of hemoglobin | Standard | Proper |
| (Blood contact Properties) | Complement activation | Activation of C3 complement | Standard | Inappropriate |
| In Vitro Immunology | Leukocyte proliferation | Leukocyte proliferation with mitogen stimulation | Standard | CCK-8 |
| (Cell-based assays) | Phagocytosis | Zymosan assay | Standard | Proper |
| | Cytokine induction | Cytokine production | Standard | Proper |
| Toxicity | Oxidative stress | Detection of ROS | Standard | Proper |
| | Cytotoxicity (necrosis) | Cell viability and mitochondrial integrity | Standard | CCK-8 |
| | Cytotoxicity (apoptosis) | Activation of caspase 3 | Standard | Annexin-V |
| Targeting | Cell binding/internalization | N/S | N/S | TEM, confocal microscope or other methods |

Characteristics specific to metal nanomaterials

- Nanoparticles larger than 100 nm tend to aggregate relatively quickly *in vitro* when compared to nanoparticles smaller than 100 nm. Fresh samples within two weeks after synthesis is recommended for tests.
- Each standard toxicology method must be verified before use. (ex. interference with a specific wavelength, electrophoresis)

Flow chart for nanotoxicity tests



Biological reactivity of silver nanoparticles

Cytotoxicity of silver nanoparticles





Cytotoxicity of silver nanoparticles



U937 cells (macrophage)

Induction of apoptosis by silver nanoparticles

20 nm

80 nm



- : U937 cells (macrophage)
- : 25 μ g/mL for 15 hrs

Lysosomal aggregation by silver nanoparticles



- : U937 cells (macrophage)
- : 20 nm, 25 $\mu g/mL$ for 24 hrs

ROS production by silver nanoparticles



Cytokine production by silver nanoparticles

• Cytokine array



Positive: chemokines (IL-8, MIF, RANTES), Serpin E1, IL-16 Negative: TNF- α , IL-6, IL-1

Cytokine production by silver nanoparticles

• ELISA (IL-8)



: U937 cells (macrophages) : 20 nm for 24 hrs Activation of signaling molecule by silver nanoparticles

• MAP kinase (ex. ERK) activation



- : Protein 30 µg loading
- : LPS (E. coli lipopolysaccharide) 50 ng/mL
- : 5 nm silver nanoparticles, 1.5 μ g/mL

Summary

- In human cells, epithelial cells from skin or lung, and macrophages, 5 nm and 20 nm silver particles induced stronger cytotoxicity and ROS synthesis than 80 nm particles did.
- 5 nm and 20 nm silver particles induced chemokine production, mainly IL-8, MIF and RANTES, while proinflammatory cytokines, IL-1, IL-6 and TNF-α were not induced significantly in the same conditions.
- Some MAP kinase signaling pathways were activated during exposure to silver nanoparticles at lower concentrations which do not induce cytotoxicity.

Conclusion

- The toxicity and inflammatory effects of nanoparticles are dependent on their size. In silver nanoparticles smaller than 20 nm induce cytotoxicity significantly *in vitro*.
- Nanoparticles induce inflammatory immune responses at lower concentrations and chemokines are the major cytokines induced at early stages of exposure to silver nanoparticles.





