

MEMS for Bio Applications

October 14, 2003

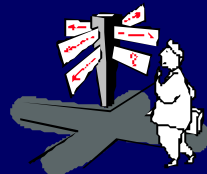
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- Bio Applications
 - Micro Array: Peptide Micro Array
 - Bead Affinity Chromatography Chips
 - Enzyme Reaction in a Micro Fluidic System
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- Summary



Applications of MEMS

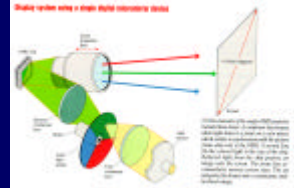
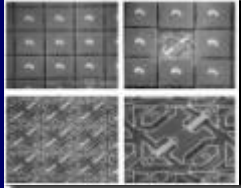
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Applications of MEMS

- MEMS has been developed as a break-through technology.



- Pressure sensors
- Accelerometer
- Gyroscope
- Digital Micromirror Device
- Inkjet Head
- Optical Switch


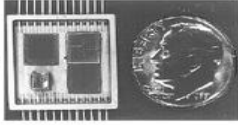


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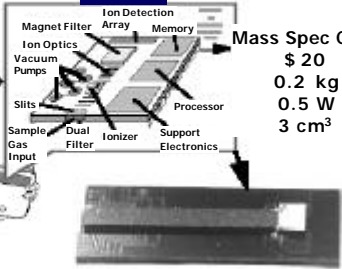
Conventional vs. MEMS Inertial Measurement Units

Conventional	MEMS
	
<p>Mass : 1587.5 grams Size : 15 cm x 8 cm x 5 cm Power : 35 W Survivability : 35 G's Cost : \$ 20,000</p>	<p>Inertial Measurement Unit</p> <p>Mass : 10 grams Size : 2 cm x 2 cm x 0.5 cm Power : ~ 1 mW Survivability : 100 kG's Cost : \$ 500</p>

- The merits of MEMS are to be small, light, cheap, multi-functional and integrated with mechanical and electronic components.

From DARPA

Mass Spectrograph on a Chip

Conventional	MEMS
	
<p>\$ 17,000 70 kg 1,200 W 20,000 cm³</p>	<p>Mass Spec Chip \$ 20 0.2 kg 0.5 W 3 cm³</p>

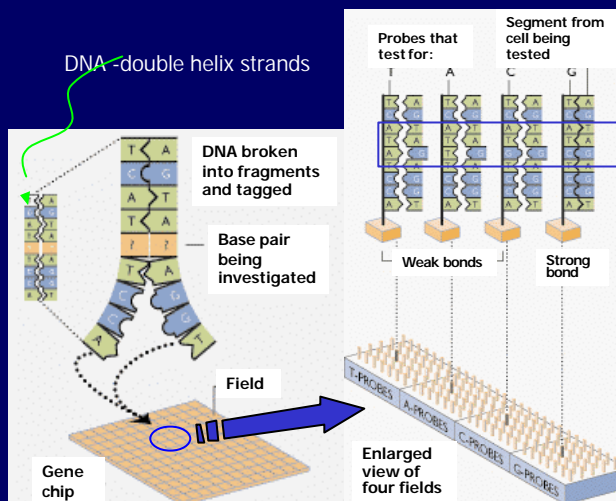
From DARPA

- Mass spectrograph on a chip will be integrated with vacuum pumps, ionizer, an ion detector array, and control electronic circuits.
- The MS on a chip enables portable measurement units and point-of-care will be available.
- BioMEMS provides new methodologies to the biotechnology.
- For examples, lab-on-a-chips, Affymetrix DNA chips, and high throughput screening chips continue to evolve the biotechnology.

Bio Applications : Peptide Micro Array Synthesized using Micromirror Array

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DNA Micro Array



- **DNA micro arrays**

- DNA sequencing assay
- Microfabrication
 - (a) Contact printing
 - (b) Inkjet printing
 - (c) In situ fabrication using inkjet
 - (d) In situ fabrication using photolithography

- **Merits**

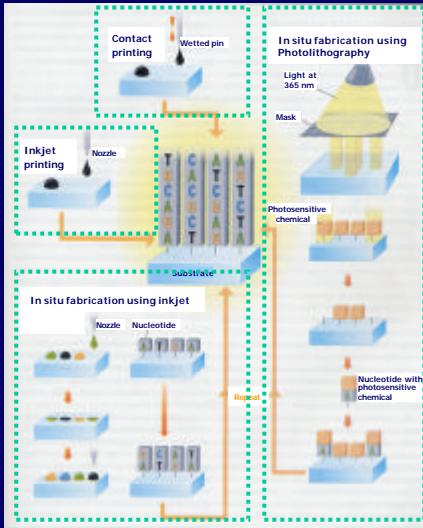
- One chip assay
- Performance is improved
- Assay time and cost are reduced
- Easy manipulation

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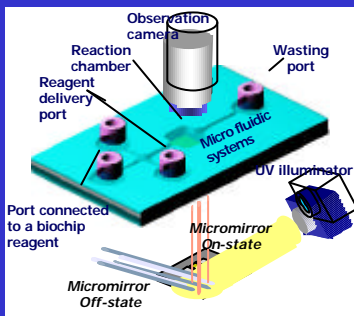
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Making Micro Arrays

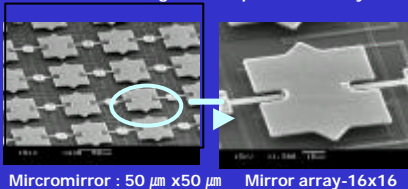


- **Making micro arrays**
 - Just printing
 - (a) Contact printing
 - (b) Inkjet printing
 - In situ fabrication
 - (c) In situ fabrication using inkjet
 - (d) In situ fabrication using photolithography
- **In situ fabrication using photolithography**
 - Light at 365nm is shone through a mask.
 - The light releases the capping chemical, exposing parts of the substrate.
 - A solution is then washed over the substrate.
 - The nucleotides attach to the unprotected sites, adding their own capping layer.
 - The process is repeated, building up sequences of DNA.
- **Suitable to mass production with the same sequence.**

Synthesis of Peptide Micro Array using Micromirror Array

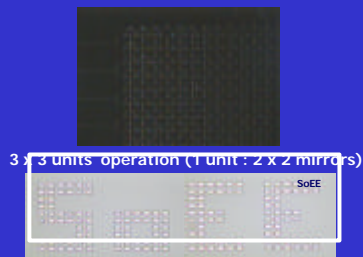


Schematic drawing of biochip fabrication system



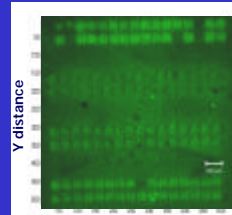
Mircromirror : 50 μm x50 μm

Mirror array-16x16



3 x 3 units operation (1 unit : 2 x 2 mirrors)

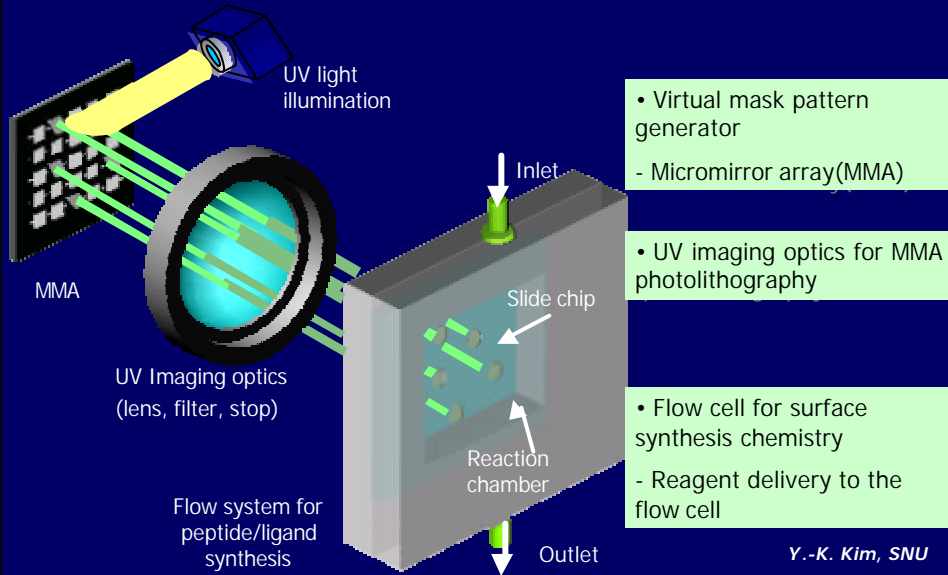
Photoresist(AZ1512) patterns produced by micromirror array



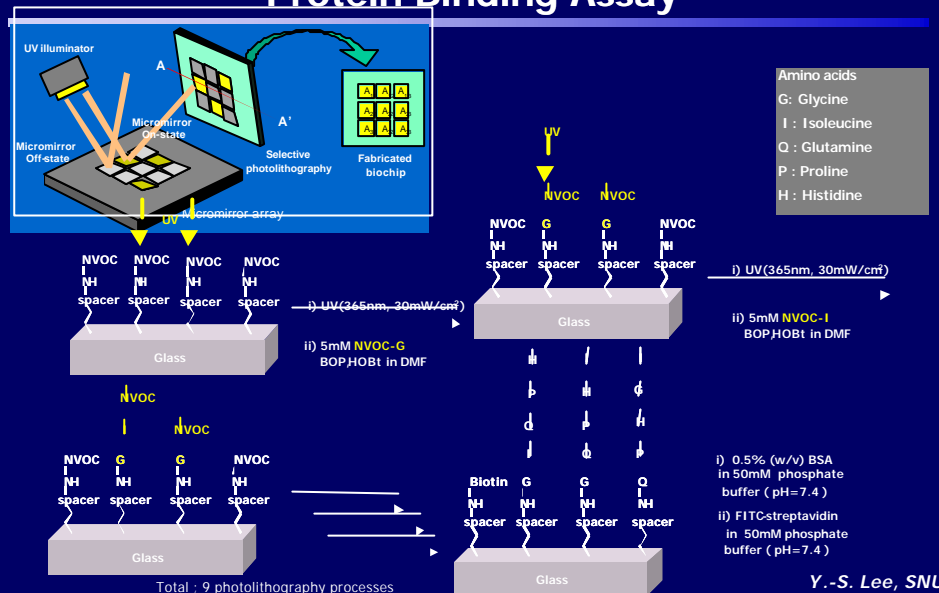
Line 1 (Biotin)
Line 2 (HPQIG)
Line 3 (IHPQG)
Line 4 (IGHPO)

Peptide micro array

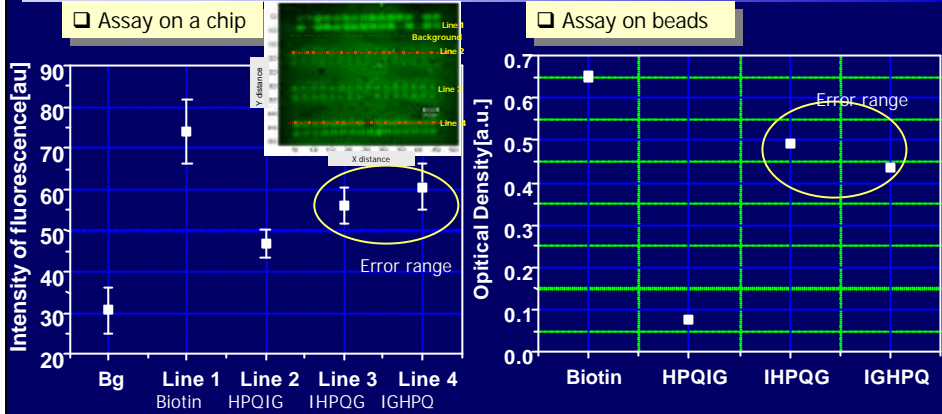
MMA Systems for Peptide Synthesis on a Chip



Peptide Array Synthesis on a Chip using MMA and Protein Binding Assay



Ligand Interaction using MMA



- Fluorescence assay vs Enzyme assay
- Sensitivity & S/N
- One chip analysis vs 4 times ELISA with sophisticated controlled condition

Assay time (labor) reduced (<1/7 days), Convenience (automated synthesis by MMA systems), Improvement of assay reliability by comparison on a chip Y.-S. Lee, SNU

Merits of the Synthesis using MMA

	On resin (bead)	On surface (chip)	Comments
Surface density	0.1 mmol/g	1 nmol/cm ²	
Quantity	resin 500 mg	5 mM solution in 200 μl flow cell	
Required amino acid	150 μmol / reaction	1 μmol / reaction	
Application example	 GIQPH QPHGI	 GIQPH GOPHI	100 mg resins for each penta peptide for 3 kinds of sequence G: Glycine, I: Isoleucine, Q: Glutamine, P: Proline, H: Histidine
Reagent quantity (Amino acids)	15 times x 150 μmol = 2250 μmol	15 μmol	3 sequences x penta peptide = 15 times
Assay time (labor)	~ 7 days	< 1 day	affinity assay of streptavidin

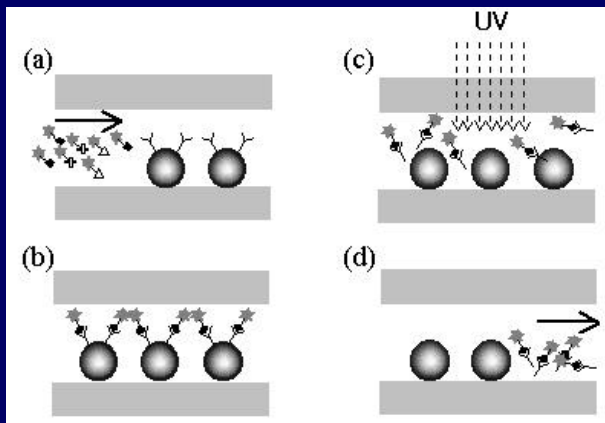
- The MMA system provides the flexibility on biopattern design and fast turn-around analysis.

Bio Applications : Bead Affinity Chromatography Chips

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Bead Affinity Chromatography (BAC)

- BAC chips are based on the idea using a large surface area of beads to analyze bio target molecules in a low concentration.
- Silicon and glass based micromachining makes the reaction chamber packed micro beads and micro channels.



- (a) Injection biosamples
- (b) Concentration & separation of target protein
- (c) Elution of target protein using UV cleavage
- (d) Analysis of target protein using mass spectrometry or fluorescence

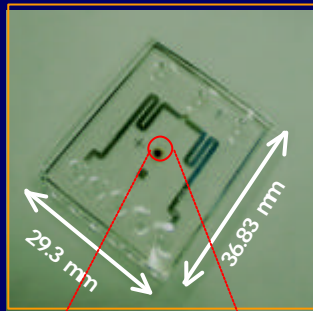
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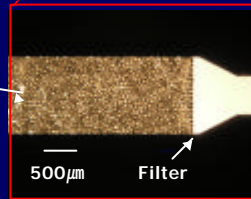
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Micro Bead Affinity Chromatography Chips



Micro Beads

Left : magnetic bead
Right : CutiCore bead



Reactor depth 50 μm
Total volume : 1 μl

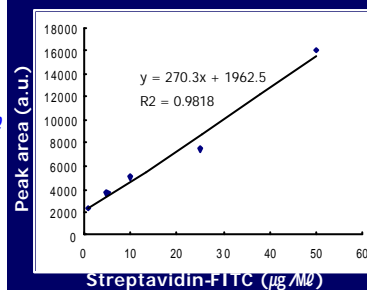
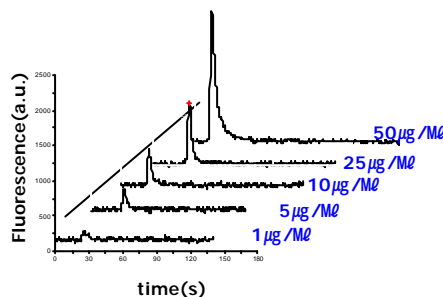
Reactor depth 100 μm
Total volume : 0.4 μl

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Quantitative Analysis Streptavidin-Biotin Model System

- Packed bead (40 μl of 0.33 % bead suspension), BSA blocking (20 min),
- Sample injection and reaction (30 min), room temperature
- Streptavidin-FITC (1 - 50 μg/Ml) / PB (pH 7.4)
- UV irradiation for 30 min and elution with PB

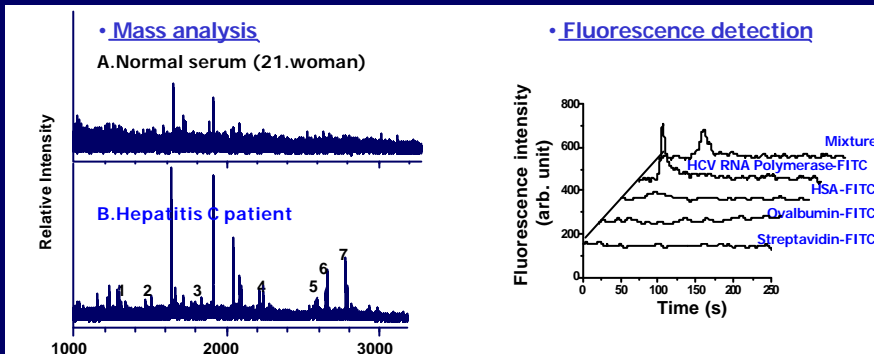
Chromatogram of eluted protein



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HCV RNA Polymerase Detection and Analysis by MALDI-TOF & Fluorescence

- To show the feasibility, we tested the HCV RNA polymerase detection.
- With MALDI and fluorescence detection, HCV RNA polymerase are detected, even in cocktail samples.



1. 1311.961(LPINALSNLLR)
2. 1511.846(YLPNWAIVKTKLK)
3. 1835.717(WIYVLLLFLLADAR)
4. 2234.986(LNAACNIVTRGERCDLEDR)
5. 2585.975(SLTERLYGGPLTNSKGNCGYR)
6. 2651.06(CRASGVLTTSCGNLTCLYKASAAQR)
7. 2779.324(ALDCQIYGACYSIEPLDLPQIER)

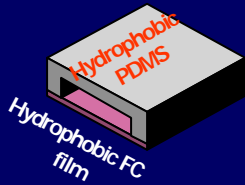
❖ 100 - 200 fmol/ μ l of HCV RNA polymerase was detected

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Bio Applications : Enzyme Reaction in a Micro Fluidic System

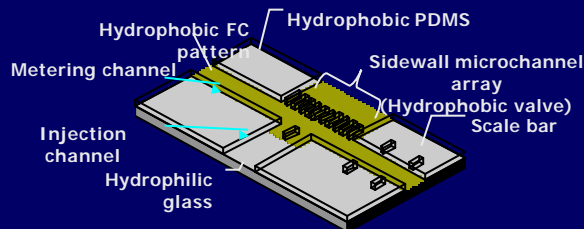
Variable Metering using a T-shaped Microinjector with Sidewall Microchannel Array

- Nanoliter droplets are controlled using the surface property such as hydrophobic and hydrophilic.

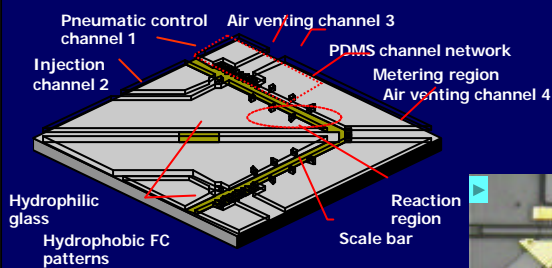


$$P = g \cos \theta_{PDMS} \left(\frac{2h+w}{hw} \right) + g \cos \theta_{FC} \left(\frac{w}{hw} \right)$$

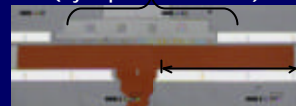
A negative pressure indicates a repellent force at the solid-liquid-gas interface.



Enzyme Reaction in a Micro Fluidic System

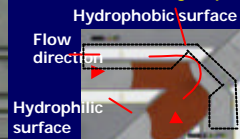


Sidewall microchannel array (Hydrophobic valve "ON")



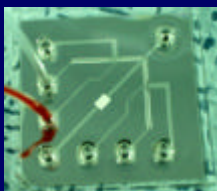
Effective length (30 nl)

• Nanoliter measuring step

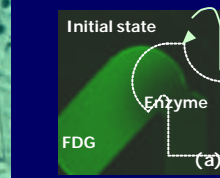


• Flow direction control step

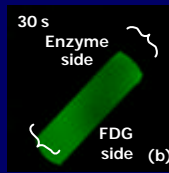
7 min 30 s



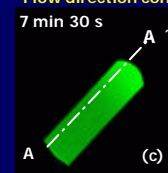
Fabricated device



fluorescein di-β-galactopyranoside



Reaction on a micro fluidic system



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Bio Applications : Measurements using Optical MEMS Components

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Bio Measurements using Optical MEMS

Schematic drawing of Bio Measurements

Labels in schematic: Fiber fixer & micro lens, Scanning mirror, Location of protein chip, Collimating lens, Micro prism, Photodetector.

Micro scanning mirror
Micro spring

2D micro scanner
Mirror : 3.5 mm x 3.5 mm

Laser spot (off state)

Scanning line (driving mirror only)

Scanning line (driving movable frame only)

Resonance of mirror at 380Hz

Resonance of movable frame at 150Hz

1. 2D surface scan (silicon spring)
2. 1:2 freq. ratio (silicon spring)
3. 1:2 freq. ratio (polymer spring)
4. 2:5 freq. ratio (polymer spring)

Lissajous pattern illuminated using micro scanner

Micro prism (PMMA) fabricated using hot embossing

Micro lens (PMMA)

Labels in lens image: PMMA, Glass

Y.-K. Kim and H. -S. Kim

Summaries

- MEMS technology provides the strong methodologies to biotechnology, life science and chemical engineering.
- Nanotechnology will also be a break-through technology to biotechnology.
- Acknowledgements
- These researches have been supported by MOST (Ministry of Science and Technology) and KOSEF (Korea Science and Engineering Foundation).
- Co workers
- Prof. Yoon-Sik Lee, Prof. Byung-Gee Kim (SNU), Prof. Hoseong Kim (ChungAng Univ.)
- Post Doc. Researchers : Dr. Kooknyung Lee, Dr. Sangho Lee, Dr. Chang-Soo Lee (SNU).
- Graduate Students: Minsoo Kim, Suhjung Cho, Woojae chung (SNU), Yongyoon Kim (ChungAng Univ.).